



CEFood

Congress

Novi Sad, Serbia  
23 - 26 May, 2012

# PROCEEDINGS



International Union  
of Food Science and  
Technology

European Association  
of Food Science and  
Technology



Central  
European  
Initiative

ISBN 978-86-7994-027-8

6TH CENTRAL EUROPEAN CONGRESS ON FOOD, Novi Sad 2012, SERBIA

***Publisher***

University of Novi Sad, Institute of Food Technology  
Bulevar cara Lazara 1.  
21000 Novi Sad, Serbia

***Main editor***

Dr. Jovanka Lević

***Editors***

Prof. Dr. Viktor Nedović  
Dr. Nebojša Ilić  
Dr. Vesna Tumbas  
Ana Kalušević, dipl. ing.

***Abstract/Paper Review***

All abstracts and papers are reviewed by the International Board of Reviewers

***Technical editors***

Bojana Kokić  
Miona Belović  
Dubravka Jambrec  
Nataša Nedeljković  
Olivera Đuragić  
Tanja Radusin  
Ana Kalušević  
Tamara Dapčević  
Tatjana Tasić  
Jovana Vučković  
Tamara Sarafijanović

***Cover***

Boris Bartula, BIS, Novi Sad, Serbia

***Printed by***

"Futura" – Novi Sad, Serbia

***Number of copies***

600 copies

## CONGRESS PRESIDENT

Prof. Dr. Viktor Nedović, Faculty of Agriculture, University of Belgrade, Serbia

## INTERNATIONAL SCIENTIFIC COMMITTEE

Prof. Dr. Peter Raspor, Biotechnical Faculty, University of Ljubljana, Slovenia

Prof. Dr. Roger Fenwick, Institute of Food Research, Norwich, United Kingdom

Prof. Dr. Dietrich Knorr, Berlin University of Technology, Germany

Prof. Dr. Brian Mckenna, University College Dublin, Ireland

Prof. Dr. Viktor Nedović, Faculty of Agriculture, University of Belgrade, Serbia;

Dr. Jovanka Lević, Institute of Food Technology in Novi Sad, Serbia

Prof. Dr. Gustavo V. Barbosa-Cánovas, Center For Nonthermal Processing of Food, Washington State University, Usa

Prof. Dr. José Aguilera, Catholic University of Chile, Chile

Prof. Dr. Eyal Shimoni, Department Of Biotechnology And Food Engineering, Technion – Israel Institute of Technology, Israel

Dr. Nebojša Ilić, Institute of Food Technology in Novi Sad, Serbia

Dr. Vesna Tumbas, Faculty of Technology, University of Novi Sad, Serbia

Prof. Dr. Branko Bugarski, Faculty of Technology and Metallurgy, University of Belgrade, Serbia

Dr. Juan Valverde, Teagasc, Ireland

Prof. Dr. Laura Piazza, Department of Food Science and Technology, State University of Milan, Italy

Prof. Dr. Taoukis Petros, School of Chemical Engineering, National Technical University of Athens, Greece

Dr. Anamarija Mandić, Institute of Food Technology in Novi Sad, Serbia

Dr. Aleksandra Mišan, Institute of Food Technology in Novi Sad, Serbia

Dr. Marijana Sakač, Institute of Food Technology in Novi Sad, Serbia

Prof. Dr. Slađana Šobajić, Faculty of Pharmacy, University of Belgrade, Serbia

Prof. Dr. Francesco Capozzi, Faculty of Agriculture, University of Bologna, Italy

Dr. Huub Lelieveld, Ghi Association Netherlands and Effost Executive Committee, Netherlands

Prof. Dr. Dominique Bauchart, INRA, Clermont Ferrand, France

Prof. Dr. Bogdan Yegorov, Odessa National Academy of Food Technologies, Ukraine

Prof. Dr. Mark Shamtsyan, St. Petersburg State Institute of Technology, Technical University of Moscow, Russia

Prof. Dr. Jana Hajslova, Institute of Chemical Technology, Prague, Czech Republic

Prof. Dr. Giovanni Dinelli, Department of Agroenvironmental Sciences and Technologies, University of Bologna, Italy

Prof. Dr. Željko Knez, Faculty of Chemistry and Chemical Engineering, University of Maribor, Slovenia

Dr. Diego Moreno-Fernández, Spanish National Research Council, Spain

Prof. Dr. Gerhard Schleining, Boku, Vienna, Austria

Prof. Dr. Živko Nikolov, Department of Biological & Agricultural Engineering, Texas A&M University, USA

Prof. Dr. András Salgó, Faculty of Chemical and Biochemical Engineering, Budapest University of Technology and Economics, Hungary

Dr. Nastasia Belc, Institute of Food Bioresources, Bucharest, Romania

Prof. Dr. Vladimir Mrša, Faculty of Food Technology and Biotechnology, University of Zagreb, Croatia

Prof. Dr. Draženka Komes, Faculty of Food Technology and Biotechnology, University of Zagreb, Croatia

Prof. Dr. Radoslav Grujić, Faculty of Technology Zvornik, University of East Sarajevo, BIH Republic of Srpska

Prof. Dr. Vladimir Kakurinov, Veterinary Faculty, St. Kliment Ohridski University, Macedonia

Prof. Dr. Vural Gökmen, Food Engineering Department, Hacettepe University, Turkey

Pof. Dr. Kemal Çelik, Faculty of Agriculture, Çanakkale Onsekiz Mart University, Turkey  
Prof. Dr. Ida Leskošek Čukalović, Faculty of Agriculture, University of Belgrade, Serbia  
Prof. Dr. Spasenija Milanović, Faculty of Technology, University of Novi Sad, Serbia  
Prof. Dr. Miroslav Vrvic, Faculty of Chemistry, University of Belgrade, Serbia  
Dr. Vesna Matekalo Sverak, Institute of Meat Hygiene and Technology, Belgrade, Serbia  
Prof. Dr. Dragojlo Obradović, Faculty of Agriculture, University Of Belgrade, Serbia  
Prof. Dr. Miomir Nikšić, Faculty of Agriculture, University of Belgrade, Serbia  
Prof. Dr. Predrag Puđa, Faculty of Agriculture, University of Belgrade, Serbia  
Prof. Dr. Andreja Rajković, Faculty of Agriculture, University of Belgrade, Serbia  
Prof. Dr. Sonja Đilas, Faculty of Technology, University of Novi Sad, Serbia  
Dr. Milica Radosavljević, Maize Research Institute Zemun Polje, Serbia  
Prof. Dr. Ljiljana Petrović, Faculty of Technology, University of Novi Sad, Serbia  
Prof. Dr. Marija Škrinjar, Faculty of Technology, University of Novi Sad, Serbia  
Prof. Dr. Svetlana Živanović, Department of Food Science and Technology, The University of Tennessee, USA  
Prof. Dr. Neda Mimica Dukić, Faculty of Sciences, Novi Sad, Serbia  
Dr. Marija Bodroža Solarov, Institute of Food Technology in Novi Sad, Serbia  
Prof. Dr. Mirko Babić, Faculty of Agriculture, University of Novi Sad, Serbia  
Prof. Dr. Vera Lazić, Faculty of Technology, University of Novi Sad, Serbia

## INFLUENCE OF MYCOTOXINS IN SWINE FEED ON THE HEALTH STATUS OF PIGLETS

Prodanov-Radulović Z. Jasna\*<sup>1</sup>, Došen Đ. Radoslav<sup>1</sup>, Stojanov M. Igor<sup>1</sup>, Pušić M. Ivan<sup>1</sup>, Živkov-Baloš M. Milica<sup>1</sup>, Urošević I. Miroslav<sup>2</sup>

<sup>1</sup>Scientific Veterinary Institute „Novi Sad“, Rumenacki put 20, 21000 Novi Sad, Serbia;

<sup>2</sup>Agriculture School, Carice Milice 2, Futog, Serbia

\*Corresponding author:

Phone: +381214895313

Fax: +38121518544

E-mail address: [jasna.prodanov@gmail.com](mailto:jasna.prodanov@gmail.com)

**ABSTRACT:** Mycotoxins are secondary metabolites of fungi that can contaminate animal feeds at all stages of food production chain. Consumption of feed contaminated with mycotoxins may result in immunosuppression, which represent a factor predisposing livestock to infectious diseases. From the epidemiological point of view, it is important to note that mycotoxins may cause breakdown of active immunity and occurrence of disease even in properly vaccinated animals. The aim of the paper was to evaluate the influence of mycotoxins on the health status of piglets. The material for this research included the samples from five swine farms, where health disorders in suckling and weaned piglets were detected. Depending on the specificity of each evaluated case and available material, the applied research methods included: epidemiological and clinical evaluation, pathomorphological examination, standard laboratory testing for detection the presence of aerobic and anaerobic bacteria, virological testing and microbiological feed testing, in order to examine the presence of fungi and mycotoxins by the method of thin layer chromatography. In our research the persistent presence of various infections, which react poorly or do not react on the applied antimicrobial therapy was discovered. The presence of mycotoxin in feed can be directly connected to the detected health disturbances in piglets on the examined swine farms (vulvovaginitis, pneumonia, gastroenteritis)

**Key words:** *mycotoxins, swine feed, piglets diseases*

## INTRODUCTION

Mycotoxins are secondary metabolites of mould, and so far, approximately 400 secondary metabolites with toxigenic potential produced by more than 100 moulds have been reported (Kabak et al., 2006). At the global level, it is considered that 25% of the world crop production is contaminated by mycotoxins, which may be a risk factor affecting human and animal health (Bouhet and Oswald, 2005). The issue of mycotoxins and mycotoxicoses in veterinary medicine is directly connected to the usage of mouldy and adversely storage of different types of grains (corn, wheat, barley) in animals feed (Osweiler, 2006).

In swine production in our geographic region, the most common are mycotoxicosis caused by zearalenon (F-2 toxin, ZEA), but also aflatoxins (AF), ochratoxin (OCT) and trichothecenes. (Gonzales and Rodriguez, 2008). The clinical and pathomorphological picture of mycotoxicoses in swine depend on the age and category (breeding animals, suckling and weaned piglets, fatteners) (Prodanov et al., 2009). The young animals are much more sensitive to the effects of mycotoxins comparing to adults. Mycotoxins have hepatotoxic, nephrotoxic and immunosuppressive effects (Kabak et al., 2006), which can further complicate clinical and pathomorphological picture and diagnosis of mycotoxicosis in swine. The biggest challenge with mycotoxicoses is the non-specific nature of symptoms in the affected animals. Consequently, the health disorders due to mycotoxins in the feed are difficult to diagnose (Osweiler, 2006; Prodanov et al, 2009). It has been recognised by veterinary clinicians that marked immunosuppression is observed in livestock ingesting mycotoxins at levels below those that cause overt toxicity (Oswald et al., 2005). Immunosuppressive effects of mycotoxins are of special interest and may have significant influence on the occurrence of infective diseases of pigs (Obremski et al., 2008; Prodanov-Radulović et al., 2011).

The aim of the paper was to evaluate the influence of mycotoxins on the health status of suckling and weaned piglets.

## MATERIAL AND METHODS

The material for this research included the samples from five swine farms, where health disorders in suckling and weaned piglets were detected. Depending on the specificity of each evaluated case and available material, the applied research methods included: epidemiological and clinical evaluation, pathomorphological examination, standard laboratory testing for detection the presence of aerobic and anaerobic bacteria, virological testing and microbiological feed testing, in order to examine the presence of fungi and mycotoxins by the method of thin layer chromatography.

## RESULTS AND DISCUSSION

In the first examined farm, the health disturbances in suckling piglets and weaners were registered. Clinically, the diarrhoea in suckling piglets already in the first 3 days of life after farrowing were detected. After supervision of the farm records several facts were discovered: diarrhoea occurs in the piglets of normal birth body weight, the percent of mortality is higher in animals in good body condition and on the weaning there is 30% of small piglets. Therapeutic treatment of piglets by oral and parenteral antibiotics application did not improve health problems. On the farm dams are twice vaccinated during gestation with the aim to prevent disease in piglets (diarrhoea) in the first days of life. By clinical examination the certain number of suckling piglets the clinical sign of vulvovaginitis (swelling and reddening of the vulva) were discovered. Carrying health control in the weaned piglets the diarrhoea and signs of pneumonia (cough, nasal secretion) were detected. The pathomorphological examination of the dead suckling piglets revealed lesions dominantly on the mucosal surface of the digestive tract (*Haemorrhagiae mucosae ventriculi*, *Enteritis catharralis acuta*). In dead weaners beside lesions in the digestive tract, the prominent pathological changes in lungs were discovered (*Pneumonia fibrinosa in statu hepatisationis rubrae et griseae*). By bacteriological testing on tissue samples deriving from dead animals the following bacteria was detected: *Escherichia coli haemolytica*, *Streptococcus alfa haemolyticus*, *Pasteurella sp.* Having in mind the clinical and pathological symptoms observed, especially the signs of vulvovaginitis in just farrowed piglets, a justified suspicion on the presence of mycotoxins in feed was made. Microbiological first feed for piglets testing detected 3-fold increase in the number of fungi genera *Fusarium sp.*, *Penicillium*, *Aspergillus*, *Rhizopus sp.* as compared to the level set by the regulation. Applying further laboratory testing increase of the total number of fungi in the large number of examined feeds was discovered: corn ( $887 \times 10^3$  *Aspergillus*, *Rhizopus*), piglets second feed ( $319 \times 10^3$  *Aspergillus*, *Mucor*, *Rhizopus*), feed for pregnant sows ( $123 \times 10^3$  *Penicillium*, *Fusarium*) and feed for lactating sows ( $526 \times 10^3$  *Aspergillus*, *Penicillium*, *Mucor*). The presence of mycotoxins was detected: zearalenon (ZEA) in the feed for pregnant sows (0.72 mg/kg), aflatoxin (AF) B1 in the piglets first feed (0.018 mg/kg) and ochratoxin (OCT) A in the piglets second feed (0.12 mg/kg).

On the second examined swine farm the health problems included increased incidence of clinical and pathomorphological signs of infective diseases. Analysing the existing data on the farm, the high mortality in piglets 7 days before weaning was noticed, which do not decrease after medical treatment. With the aim to overcome the problem, the measure of medical treatment of piglets 3 days before weaning was introduced, but again with no result. In the weaned piglets the disease was clinically characterised with the signs of severe yellowish diarrhoea, dehydration, huddling, roughly hair and sporadically coughing. Therapeutic treatment of the diseased animals was multiple: the antibiotics were given through feed, water and parenterally. Applying pathomorphological examination on the dead weaned piglets, the prominent changes on the digestive and respiratory tract were detected (*Gastroenterotyphlitis haemorrhagica*, *Poliserositis fibrinosa massiva*, *Pneumonia complex*).

By bacteriological testing on tissue samples from dead piglets the following bacteria were isolated: *Escherichia coli haemolytica*, *Pasteurella sp.*, *Streptococcus uberis*. By laboratory feed testing i. e. the available first feed for piglets the presence of OCT (0.5 mg/kg) and ZEA (4 mg/kg) was discovered.

The presence of mycotoxin in feed for pregnant sows have influence on the occurrence of decreased immunological defence in piglets (Prodanov et al., 2009). As a consequence of immunosuppressive action of mycotoxins, clinical and pathological lesions correspond to the infective diseases of different ethiology (Kabak et al., 2006; Obremski et al., 2008). Combinations of several mycotoxins may potentiate the action of one other, or at least exert an additive effect (Osweiler, 2006). Nutritional effects associated with feed refusal may also contribute to observed decreased efficacy of therapeutic treatments and vaccination (Oswald et al., 2005). In the second evaluated case, the presence of OCT and ZEA in the swine feed was detected. Consequently, on the farm an evident decrease in the swine immunity against infective diseases was noticed and there was no positive response on the applied antibiotic therapy. Also, the occurrence of diarrhoea and increased percent of waste piglets can be connected with presence of the mycotoxins in the feed, because the piglets display clinically feed refusal. The gastrointestinal tract represents the first barrier against ingested food contaminants and natural toxins. Following ingestion of mycotoxin contaminated feed, intestinal epithelial cells could be exposed to a high concentration of toxin (Bouhet and Oswald, 2007). From a public health perspective, increased infections in animals may result in increased animal-to-human transmission of pathogens and/or increased antibiotic concentrations in meat, as a consequence of animal treatment (Oswald et al., 2005).

The complex health problems on the third swine farm were noticed. Applying control of the anamnestic data, lately the frequent periods when sows delivery mummified piglets, stillbirths and decreased litter size were observed. Beside this, just farrowed piglets are nonviable and despite the medical treatment, they live only 3-4 days after birth. In the pregnant dams the immunoprophylaxis is carrying out with the aim to prevent the outbreak of disease in suckling piglets. Beside this, the antibiotics are added in the sows feed 7 days before and 7 days after farrowing. Sporadically, the occurrence of severe yellowish diarrhoea in piglets and apparent clinical signs of vulvovaginitis in just born piglets were evident. In the weaners the clinical signs of diarrhoea and pneumonia are sporadically noticed. The pathomorphological examination of the dead weaners revealed lesions on the organs of respiratory (*Pleuropneumonia actinobacillosa*, *Pneumonia interstitialis*) and digestive tract (*Gastroenterotyphlitis haemorrhagica acuta*). Applying bacteriological examination on the tissue samples deriving from dead piglets the following bacteria were isolated: *Escherichia coli haemolytica*, *Pasteurella sp.*, *Actinobacillus suis*. After laboratory testing of swine feed samples the simultaneous presence of several mycotoxins was established: ZEA (6.4 mg/kg), AF (0.0064 mg/kg), OCT-A (0.032 mg/kg).

For the known mycotoxins of clinical importance for swine production, the response is usually subacute or chronic and the presenting signs are often subtle and vague (Gonzales and Rodriguez, 2008). The continuous intake of small amounts of mycotoxins leads to chronic intoxication which is clinically characterized by the loss of weight, insufficient weight gain and increased susceptibility for infectious diseases (Osweiler, 2006). Ingestion of low doses of mycotoxins can increase intestinal colonization by opportunistic pathogenic bacteria in piglets (Oswald et al., 2005; Taranu et al., 2005). Mycotoxins mixtures i.e. the combinations of several mycotoxins are likely to occur naturally and they may influence on the immunity in an additive or synergistic manner. Economic losses that occur as a consequence of interaction of several mycotoxins are still unknown because in low concentrations several mycotoxins may interact in way that is difficult to detect. Combinations of several and more moderate concentrations of different mycotoxins, which individually may appear to be too low in level to be a concern, can cause cumulative toxicosis, which affect the ability of the pig's organism to fight diseases (Diekman and Green, 1992; Osweiler, 2006).

On the fourth examined farm, clinically in suckling piglets the signs of severe disturbance of the central nervous system (wide open eyes, paddling, trembling, ataxia, paresis and paralysis) were detected. In some cases the whole litter of piglets died within 48 hours.

Despite the fact that the piglets were therapeutically treated, there was no evident respond to applied medication. The pathomorphological changes that were detected in dead sucklings indicated the lesions characteristic for *Morbus Aujeszky* infection (MA) (*Necroses miliaris hepatis, Haemorrhagiae corticis renis, Tonsillitis diphtheroides necroticans*). By microbiological testing in feed for lactating sows the presence of fungi (*Fusarium sp.*, *Mucor*) and AF (0.02 mg/kg) were detected. Applying virological testing (viral isolation on the susceptible cell culture) from the tissues deriving from dead piglets the MA virus was isolated.

Aflatoxins (B1, B2, G1, G2) are recognized as immunomodulatory agents, and when AFB1 is metabolised by mammals occurs in milk as M1. It is assumed that AFB1 is the most toxic fraction (Oswelder, 2006; Živkov-Baloš et al., 2008). In the case where the outbreak of MA on the farm were examined, mycotoxin (AF) in the feed can be connected with the possible reactivation of chronic (latent) infection. It has been discovered that aflatoxins decrease resistance to bacterial, fungal, viral and parasitic diseases in swine. Subsequently, vaccinations against various infective diseases may be less effective in animals exposed to mycotoxins (Diekman and Green, 1992). Even when is present in low doses, AF alters the immune response and this may predispose pigs to infectious diseases. From the epidemiological point of view, it is important that mycotoxins may cause breakdown of active immunity and occurrence of disease even in properly vaccinated animals (Marin et al., 2002; Oswald et al., 2005; Taranu et al., 2005).

In the last examined case, in suckling piglets the occurrence of neonatal diarrhoea already in the first 3 days of life after farrowing were detected. These health problems did not improve after the medical treatment with antibiotics. In great number of just farrowed piglets the most prominent clinical sign was vulvovaginitis (swelling and reddening of the vulva). Beside this, a large number of small, weak and splayleg piglets were noticed. The newborn piglets were weak, nonviable, with diarrhoea. The diseased piglets lived only for 4 days after birth. They probably died due to hypoglycemia, because sows did not have enough milk or the piglets were too weak and did not have enough strength for milk suckling. Applying pathomorphological examination on the dead suckling piglets the prominent changes on mucosal surfaces of the digestive tract (*Gastroenteritis haemorrhagica*), less number of pale kidneys, necrotic and dystrophic processes on liver tissue were detected. By laboratory testing of the available swine feed grains, the presence of ZEA in different concentrations was detected (from 0.72 to 6.4 mg/kg). The ZEA mycotoxicosis in weaned piglets was clinically characterised with signs of pneumonia, slow growth, vulvovaginitis and necrosis of the tails, sporadically with diarrhoea and rectal prolapses. The pathomorphological examination of the dead weaners revealed the following lesions: bleeding on the mucosal surface of the digestive tract, pleuropneumonia and pneumonia, hepatomegalia, focal nephritis and rectal prolapses. Etiologically the pneumonia was caused by *Actinobacillus pleuropneumoniae*, *Haemophilus suis*, and *Mycoplasma hyopneumoniae*. Also the problem that was frequently observed was digestive infection with enteropathogenic *Escherichia coli*, which can be potentiated with the detected mycotoxin (ZEA 0.8 mg/kg) and a high number of different fungi species in weaners feed (*Fusarium*, *Penicillium*, *Aspergillus*, *Rhizopus*).

Zearalenone is a mycotoxin which as an estrogen binds competitively to estrogen receptors of the uterus, mammary gland, liver and hypothalamus (Gajecki, 2002). Pigs are the most prone to the presence and negative effects of ZEA (Diekman and Green, 1992; Obremski et al., 2003). In our research, the perinatal hyperestrogenic syndrome was a constant clinical sign in suckling piglets. This is certainly the consequence of mycotoxins presence in feed for sows, e.i. during the pregnancy and the presence of its excreted metabolite in milk of the exposed sows.

## CONCLUSIONS

The influence of mycotoxins on immune system is of special interest in swine industry. The technology on swine farms demands frequent vaccinations, especially in piglets and sows which may be a problem in the case of immunocompromised animal. From the obtained



results an example of immunosuppressive effect can be presented i.e. the occurrence of enterotoxemia in piglets, despite the fact that dams were vaccinated twice during gestation. The enterotoxemia is caused by pathogenic bacterial strains and occurs frequently as a cause of mortality in the young categories. It can be provoked with the feed quality i. e. the presence of mycotoxins.

The presence of mycotoxins in feed can be directly connected to the detected health disturbances of the examined piglets. In our research we discovered the persistent presence of various infections, which react poorly or do not react on the applied antimicrobial therapy (gastroenteritis, pneumonia). Also, the chronic disturbances, for instance slow growth, malnutrition, vulvovaginitis suggest on the potential presence of mycotoxins.

## ACKNOWLEDGEMENTS

This paper is a result of the research within the project TR 31084 "Praćenje zdravstvenog stanja divljači i uvođenje novih biotehnoloških postupaka u detekciji zaraznih i zoonoznih agenasa - analiza rizika za zdravlje ljudi, domaćih i divljih životinja i kontaminaciju životne sredine (Wild animal health monitoring and introduction of new biotechnology procedures in detection of infectious and zoonotic agents – risk analysis for human health, domestic and wild animal health and for environmental contamination)", financed by the Ministry of Education and Science, Republic of Serbia.

## REFERENCES

- 1 Bouhet, S., Oswald, I.P. (2005). The effects of mycotoxins, fungal food contaminants, on the intestinal epithelial cell-derived innate immune response. *Vet Immunol Immunopathol.* 108 (1-2), 199-209.
- 2 Bouhet, S., Oswald, I.P. (2007) The intestine as a possible target for fumonisin toxicity. *Mol Nutr Food Res.* 51(8), 925-931
- 3 Diekman, M.A., and Green, M.L. (1992) Mycotoxins and reproduction in domestic livestock. *J Anim Sci*, 70, 1615-1627
- 4 Gajecki, M. (2002). Zearalenon-undesirable substances in feed. *Pol J Vet Sci.* 5 (2), 117-122.
- 5 Gonzales, S., Rodriguez, J. (2008). Mycotoxins occurrence: Animal health & Food safety concern for food animal production. In Proceedings of the 20th IPVS Congress (p. 506), Durban, South Africa
- 6 Kabak, B., Dobson, A. D., Var. I. (2006). Strategies to Prevent Mycotoxin Contamination of Food and Animal Feed. *Critical Reviews in Food Science and Nutrition*, 46, 593-619.
- 7 Marin, D.E., Taranu, I., Bunăciu, R.P., Pascale, F., Tudor, D.S., Avram, N., Sarca, M., Cureu, I., Criste, R.D., Suta, V. and Oswald, I.P. (2002). Changes in performance, blood parameters, humoral and cellular immune responses in weanling piglets exposed to low doses of aflatoxin. *J Anim Sci*, 80, 1250-1257
- 8 Obremski, K., Gajecki, M., Zwierzchowski, W., Zielonka, L., Otrócka-Domagala, I., Rotkiewicz, T., Mikolajczyk, Gajęcka, M., Polak, M. (2003). Influence of zearalenone on reproductive system cell proliferation in gilts. *Pol J Vet Sci.* 6, 239-245
- 9 Obremski, K., Zielonka, L., Gajęcka, M., Jakimiuk, E., Bakula, T., Baranowski, M., Gajecki, M. (2008). Histological estimation of the small intestine wall after administration of feed containing deoxynivalenol, T-2 toxin and zearalenone in the pig. *Pol J Vet Sci.* 11, 339-345.
- 10 Oswald, I.P., Marin, D.E., Bouhet, S., Pinton, P., Taranu, I. & Accensi, F. (2005). Immunotoxicological risk of mycotoxins for domestic animals. *Food Additives & Contaminants.* 22(4), 354-360
- 11 Osweiler, G.D. (2006). Occurrence of Mycotoxins in Grains and Feeds. In Straw, E. B., Zimmerman, J.J., D Allaire, S., Taylor, D. J., Diseases of Swine, Vol. 1153 (pp 915-930). Ames Blackwell Publishing.
- 12 Prodanov, J., Došen, R., Pušić, I., Stojanov, I., Ratajac, R., Živkov-Baloš, M. (2009). The clinical and pathomorphological diagnosis of mycotoxicosis in different swine categories. *Proc. Nat. Sci.* 116, 281-287
- 13 Prodanov-Radulović, J., Došen, R., Stojanov, I., Pušić, I., Živkov-Baloš, M., Ratajac, R. (2011) Interaction between the mycotoxins and causative agents of swine infective diseases. *Proc. Nat. Sci.* 120, 251-259
- 14 Živkov-Baloš, M., Mihaljev, Ž., Kovačević, M., Bugarski, D. (2008) Ekskrecija aflatoksisna mlekom rizik za potrošače. *Arh. vet. med.* 1, 64-68.
- 15 Taranu, I., Marin, D.E., Bouhet, S., Pascale, F., Bailly, J., D., Miller, J.D., Pinton, P., Oswald, I.P. (2005). Mycotoxin Fumonisin B<sub>1</sub> Alters the Cytokine Profile and Decreases the Vaccinal Antibody Titer in Pigs. *Toxicological Sciences.* 84, 301-307.